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## Direct culture of gonococci in the patient care area

Suspected gonococcal cervicitis or urethritis is usually confirmed by culture. Direct culture onto a selective medium in the patient care area followed by immediate incubation of the plates at the right atmosphere is considered to be the most sensitive method in the diagnosis of gonococcal infections.1 If direct plating is not possible, the samples may be either transported in nonnutritive holding media such as Stuart or Amies medium, or in nutritive transport systems. We tested the ability of a commercial nutritive transport system, Biocult-CCR (Orion Diagnostica, Espoo, Finland) to support the survival and growth of gonococci.

The urethral and cervical samples were taken at the Department of Sexually Transmitted Diseases (STD), Aurora Hospital, Helsinki, by two STD specialists, and inoculated directly at the patient's side first onto a modified Thaver-Martin (TM) medium (BBL GCII) agar base with 1.7% of haemin, 5% chocolated horse blood, 1% Iso-Vitalex, vancomysin 1 mg/l, mycostatin 50 × 105 IU/l, colimycin 0.6 mg/l, trimethoprim 2 mg/l and clindamycin 0.2 mg/l) and next, using the same swab, onto the agar surfaces of the Biocult-GC transport system. The TM plates were immediately placed in a 35°C incubator in a humidified atmosphere of 5% CO2 in air. The Biocult-GC transport device was transferred into its container, CO2 generating tablet added, the container closed tightly and placed at 35°C. The TM plates were transferred to the microbiology laboratory on the next floor the following morning, and Biocult-GC transported to another microbiology laboratory after 15 to 48 h incubation at the STD Department. The TM plates and Biocult-GC were examined for growth of gonococci after 48 hours of culture by two independent microbiologists. N gonorrhoeae growing on TM plates were identified by colony morphology, Gram stain, positive oxidase test, agglutination by monoclonal antibody (Phadebact Monoclonal GC Test 50, KaroBio Diagnostics Ab, Huddinge, Sweden), and production of acid from glucose but not from maltose or lactose. The sensitivity to penicillin (by E-test<sup>R</sup>, Biotest, Stockholm, Sweden) and ciprofloxacin (by diffusion test, Rosco, Taastrup, Denmark) as well as beta-lactamase production were also determined. N gonorrhoeae on Biocult-GC were likewise identified by colony morphology, Gram stain, oxidase positivity and growth characteristics.

First, 199 consecutive samples were studied. Smears of four patients revealed intracellular diplococci in methylene blue staining. Growth of N gonorrhoeae was detected in three samples (1.5%) on TM plates and on Biocult-GC (Table, group 1). One of the samples grew only on TM plates, another on Biocult-GC only.

In the second part of the study, samples from patients with clinically suspected N gonorrhoeae infection and from two patients not suspected to have gonococcal infection but visiting the STD Department on the same day as the index patient were studied. Intracellular diplococci were found in 33 smears from the 102 patients. N gonorrhoeae was detected by culture in 33 samples from 30 patients. Cultures from 29 patients were positive on TM plates and from 26 on Biocult-GC (Table 1, group II). Biocult-GC failed to show growth of N gonorrhoeae in samples of four patients growing on TM and TM from one growing on Biocult-GC. The difference between the yields on TM and Biocult-GC was statistically not signficant.

Although the overall agreement between TM and Biocult-GC cultures was good, five samples failed to grow on Biocult-GC and two on TM. This result may be biased as Biocult-GC was always inoculated after the TM plate (but using the same swab for both), and might have had a smaller inoculum than TM.

Gonococci are fastidious organisms and may survive in nonnutritive transport tubes for less than 24 hours. In this study Biocult-GC supported the survival and growth of N gonorrhoeae to the same extent as direct plating onto TM plates in the patient care area and may thus be an alternative to nonnutritive transport tubes if direct inoculation of samples onto TM plates in the patient care area is not possible, but inoculation, incubation and safe transport of cultured Biocult GC is. Presumptive identification of

Table Detection of N. gonorrhoeae on Thayer-Martin (TM) plates and Biocult-GCR

Group	n	Growth of gonococci					
		TM	Biocult-GC	Total	₽ <sup>†</sup>		
I.	199 (153)*	3 (3)	3 (3)	4 (4)	0		
II AII	131 (102) 330 (255)	31 (29) 34 (32)	28 (26) 31 (29)	33 (30) 37 (34)	0·1969 0·1536		

66%~(102) of the patients were females in group 1, and 43% (43) in group II.

<sup>†</sup>Yield on Thayer-Martin plates and Biocult-GC compared by Chi square test; all p-values not significant.

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N gonorrhoeae (by colony morphology, Gram stain, oxidase positivity, and agglutination by monoclonal antibody) would be available from both Biocult-GC and TM plates inoculated at patient care area in 24 to 48 hours, and final results with antibiotic sensitivity 24 hours later. Savings in labour costs and culture materials may also be achieved as only Biocult-GC and TM plates with colonies suspected to be N gonorrhoeae need to be further studied and cultured at the microbiological laboratories. It has to be emphasised, however, that our results were obtained in a department where the samples were taken by STD specialists, and in a department which is supported by a medical microbiology laboratory next door. A comparison between Biocult-GC nutritive transport system and nonnutritive transport media should be performed in primary health care centres in order to test the survival of gonococci in the two systems.

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## Hepatitis B infection post Engerix B vaccination

We report a case of documented hepatitis B virus (HBV) infection despite active immunisation with Engerix B vaccine and positive hepatitis B surface antibody (anti-HBs).

A 21 year old homosexual man was first diagnosed to be HIV antibody positive in January 1991. He remains asymptomatic. His latest T helper cell count in January 1993 was 0.42 × 10°/l. Hepatitis serology in January 1991 showed negative hepatitis B surface antigen (HBsAg) and negative hepatitis B core antibody (anti-HBc), both performed with radioimmunoassay (RIA).

Three doses of 20  $\mu$ g (1 ml) of Engerix B (Smith Kleine French) hepatitis B vaccine were given by deep intramuscular injection in March, May and October of 1991. Three months later he still showed negative HBsAg, antiHBc and anti HBs (measured by enhanced luminescent assay). A fourth dose

of Engerix B was given in April 1992. Two months later he showed negative HBsAg, negative anti-HBc, but weakly positive anti-HBs, with a titre of only 14 mIU/ml. A further dose of vaccination was therefore recommended.

However, the patient defaulted follow-up for the next six months and returned in January 1993 for a routine check-up. The fifth dose of Energix B vaccine was then given.

Repeat hepatitis serology two weeks later showed positive HBsAg, positive anti-HBc, positive HBeAg, negative anti-HBs, negative anti-HBe and negative anti-HBc IgM. His asparate amino transferase (AST) level was elevated to 290 U/l (5-35 U/l). Nine days later he had negative HBsAg, positive anti-HBc IgM, negative HBeAg and positive anti-HBe. His AST was 91U/l. He remained totally asymptomatic during this period, and had a normal serum bilirubin level (11  $\mu$ mol/l). These results indicate that he had a subclinical hepatitis B infection (table). He admitted to unprotected penetrative sexual intercourse with his regular partner, whose HIV and hepatitis status is not known. He is not known to be an intravenous drug user or have any other risk factors for hepatitis B infection.

This case illustrates several interesting points. Firstly, the time required for effective hepatitis immunisation can be quite protracted, especially in the initial non-responders. Patients can therefore become infected with hepatitis B virus during the process of immunisation.

Secondly, anti-HBs level of 14 miu/ml failed to protect this patient from hepatitis B infection, although it may have served to protect him against clinical disease and against becoming a chronic carrier. HBV infection events are known to occur in subjects who are either poor or non-responders to vaccine.<sup>12</sup> The highest anti-HBs titre described which failed to protect against HBV infection was 23 samples to ratio units, which are equivalent to our miu/ml.<sup>1</sup> In our laboratory, we consider anti-HBs level below 100 miu/ml as poor responders and recommended booster doses for these patients.

Thirdly, this patient's hepatitis infection was detected through a laboratory screening programme which included HBsAg and anti-HBc as the primary tests. Anti-HBs is included for those with a history of vaccination or those who are anti-HBc positive, indicating natural infection in the past. This case would not have been discovered if testing were confined to looking for anti-HBs. It is

Table Hepatitis serology and AST results

Date	HBsAg	Anti HBc Total	Anti HBc IgM	HBeAg	Anti HBe	Anti HBs	AST U/l
24.01.91	_	_					
14.01.92	_	_				0	
10.03.92	_	_				ŏ	
10.06.92	_	_				14miu/ml	
27.01.93	+	+	_	+		0	290
04.02.93	_	+	+	<u>-</u>		•	91